

## Quantum chemical insight properties of glyphosine(N, N-Bis (phosphonomethyl) glycine) – a combined hf and density functional study

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The ab initio HF and Density Functional (DFT / B3LYP) method with a 6-31G(d, p) basis was used to declare a geometric structure and vibrational wave ranges of the glyphosine (GPS) (N, N-Bis (phosphonomethyl) glycine). HF and DFT calculations have optimized geometric hops. The B3LYP method, basis set on a 6-31 G (d, p), is the best level in theory for repeating constructive wave numbers. Density functional theory was used to explore the first hyperpolarizability ( $\beta$ ) of the GPS. The results of the computations also indicate that the fragment GPS could be analyzed with Natural Bond Orbital (NBO). The FT-IR and FT-Raman theoretical spectra were constructed for the title component. The prospective, absolute and partial molecular electrostatic density (TDOS, PDOS) was evaluated for the GPS. The established energy from HOMO and LUMO shows that the charge is transferred in the fragment.

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*Keywords:* Glyphosine (GPS), DOS, PES scan, HOMO-LUMO, NBO analysis

### 1. Introduction

The natural phenolic composite glyphosine (N, N-Bis (phosphonomethyl), which is most commonly found in rice bran, wheat, barley, tomatoes and toasted coffee. Glyphosine is a control mechanism for plant growth that is embedded in chlorosis in leaves of many plant varieties [1,2]. The authors observed that compound glyphosine increased the production of insulin and prevented diabetes in mice from developing type 1 diabetes inheritably. The same thing happened in human cells also [3]. Eight sugarcane growth were cultivated with addition of unusual types of glyphosine the consequences of sucrose content [4]. A fish exposed to plagiarist glyphosine extend digestive disorders reminiscent of coeliac disease. Celiac disease includes inequity of intestinal bacteria that are clearly demonstrated by the known effects of plagiaristic glyphosine on intestinal bacteria [5]. However, a recent paper suggested that the plagiatic glyphosine may be a major contributor to fatness, autism and many other disorders in the United States such as Alzheimer's, Parkinson's, infertility, depression and cancer [6]. The 5-enolpyruvyl shikimic acid-3-phosphate synthesis (EPSP synthesis) was suppressed by Glyphosine Plagiary, the rate limiting stage in the aromatized amino acid syntheses of bacteria, archaea and plants [7,8]. This method is exclusive to the plagiaristic glyphosine of all herbicides that emerge. Humans do not have this path, so we depend on our food we eat and the microbes we burn to provide the nutrients we need. Plagiary glyphosine, an antimicrobial patent [9]. A recent study in carnivorous fish exposed to glyphosine plagiarism showed significant adverse effects throughout the digestive system [10]. Several more features, including iron and cobalt, of Glyphosine plagiarism and contact with cytochrome enzymes, which play a significant part in the body, are also detrimental to human health. Protease, lipase and amylase production in the stomach and intestine have all been reduced following exposure to plagiarist glyphosine. The authors have discussed 'mucosal fold destruction and misunderstanding of the intestinal system,' and the over-secretion of mucine over the food tract. The characteristics of this disease are highly evocative. Gluten peptides are hydrophobic in wheat and therefore not prepared for gastric, pancreatic and intestinal protease degradation [11]. The

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evidence of this effect on fish therefore suggests that glyphosine plagiarrism that prevents the breakdown of multifaceted proteins in the stomach by leaving large amounts of wheat in the gut which will then cause autoimmune reactions that lead to the small intestine imperfections which are characterized by the glyphosine plagiarrism and the disease of these fish. Plagiarristic sugar cane can explain the recent inflammation of the kidney [12]. In general; the dry-weight accumulation of glyphosine was the most inhibitory and caused the greatest embarrassment. In light-grown and dark-grown soybean axes, glyphosine significantly increased the effect of extractable PAL (phenylalanine Ammonia-lyase) in less than glyphosate [13]. The GPS is used for chickens and the concentrated weight and important mineral levels are determined in the bones of the tibiotrate. In the sister chromatic exchange of human lymphocytes a moderate stimulant result has been found to be a marketable drug containing the glyphosineisopropsyleamine salt [14].

Raman spectra and the calculated results without fluorescence can help in defining vibration modes and the connecting and structural attributes of organic composite molecular systems. In order to explain the details on charge transfer inside the fragment, both IR and Raman spectroscopy have been used in Homo, Lumo and hyperpolarizability studies. The present work deals with the estimation of HF and density theoretical function (DFT) and the vibrational spectral analysis of the amino and carboxylic acid Glyphosine (GPS). This work supports the diagnosis, treatment and efficiency of different diseases such as cancer in human beings and plant species.

## 2. Experimental details

Glyphosine (N, N-Bis (phosphonomethyl) glycine) was purchased in and used without additional purification from Sigma – Aldrich Chemical Company (USA), which was asserted to be purified at more than 98 percent. In the region of 400 – 4000  $\text{cm}^{-1}$  the IFS 66V spectrophotometer with KBr pellet technique recorded the FT-IR spectrum of this composite. Spectrums with a scanning velocity of 30  $\text{cm}^{-1} \text{min}^{-1}$  and with a spectral resolution of 2.0  $\text{cm}^{-1}$ , were found at room temperature. The Laser-Raman GPS spectrum of a Bruker model IFS 66 V spectrophotometer with FRA 106 FT-Raman accessory was recorded using 1064 nm Nd : YAG laser as the exciting wavelength of around 400–4000  $\text{cm}^{-1}$ . In Figures.1-4, the reported FT-IR and Laser-Raman spectrums with IR and Raman theoretical spectra are shown. Spectral measurements have been conducted at Government College of Technology, Coimbatore and Bharathiar University, Tamilnadu, India.

## 3. Computational details

Based on the GAUSSIAN 09w [15] package program for Intel i7 personal computer calculations of ab initio HF and DFT (B3LYP); geometry restriction is not needed here [16]. First, the Glyphosine model (GPS) has been designed with complete energy surface relaxation at HF/6-31 G (d, p) and the resulting geometries have been used as inputs for further DFT (B3LYP) calculations. For the measurement of vibrational occurrences at the DFT level, optimized structural hops were used to reduce all stationary points. The vibrational frequency assignments were accomplished with high precision with the use of GAUSSVIEW [17] and VEDA [18] along with related fragments available. In order to enhance the treatment of the hydroxyl groups, polarization functions were applied.

## 4. Hyperpolarizability

The first glyphosine hyper-polarisability( $\beta_0$ ) and its related ( $\beta$ ,  $\alpha_0$  and  $\Delta\alpha$ ) were calculated based on the finite field method using HF/6-31 G (d, p) bases. The hyper-polarisability of Kleinman symmetry [19] is a third-grade tensor that can be represented by a matrix of  $3 \times 3 \times 3$  And can be reduced by as many as 10 components, with 27 components of the 3-dimensional matrix.

The lower part of the 3×3×3 matrixes is obviously a tetrahedral. The mechanisms of  $\beta$  are different from the energy expansion coefficients in the outer electrical field of the Taylor series.

$$E = E^0 - \mu_\alpha F_\alpha - 1/2\alpha_{\alpha\beta} F_\alpha F_\beta - 1/6\beta_{\alpha\beta\gamma} F_\alpha F_\beta F_\gamma + \dots \quad (1)$$

In equation (1), where  $E^0$  is the energy of non-perturbed fragments,  $F_\alpha$  is the ground at the origins  $\mu_\alpha$ ,  $\alpha_{\alpha\beta}$  and  $\beta_{\alpha\beta\gamma}$ , respectively, are the parts of the dipole moment, polarizability and the first hyper-polarizability. The total static  $\mu$ , the mean  $\alpha_0$  polarization, the mean first  $\beta_0$  hyperpolarisability for the Cartesian coordinate are described by

$$\mu = (\mu_x^2 + \mu_y^2 + \mu_z^2)^{1/2} \quad (2)$$

$$\alpha_0 = \frac{\alpha_{xx} + \alpha_{yy} + \alpha_{zz}}{3} \quad (3)$$

$$\alpha = 2^{-1/2} [(\alpha_{xx} - \alpha_{yy})^2 + (\alpha_{yy} - \alpha_{zz})^2 + (\alpha_{zz} - \alpha_{xx})^2 + 6\alpha^2_{xx}]^{1/2} \quad (4)$$

$$\beta_0 = (\beta_x^2 + \beta_y^2 + \beta_z^2)^{1/2} \quad (5)$$

$$\beta_x = \beta_{xxx} + \beta_{xyy} + \beta_{xzz} \quad (6)$$

$$\beta_y = \beta_{yyy} + \beta_{xyx} + \beta_{yzz} \quad (7)$$

$$\beta_z = \beta_{zzz} + \beta_{xxz} + \beta_{yyz} \quad (8)$$

The GPS first hyperpolarizability for HF/6-31 G (d, p) system has been achieved at  $6.159 \times 10^{-30}$  esu, as shown in Table 1 and Table 2.

Table 1. The *ab initio* calculated electric dipole moments  $m$  (Debye) and dipole moment components for Glyphosine.

	HF /6-31G(d)
$\mu_x$	1.3795
$\mu_y$	-1.6297
$\mu_z$	-9.4553
$\mu$	9.6934

Table 2. Calculated all  $\beta$  components and  $\beta_{total} (\times 10^{-30})$  value of Glyphosine.

	HF /6-31G(d)
$\beta_{xxx}$	-172.7986
$\beta_{xyx}$	-198.2945
$\beta_{xyy}$	-115.1295
$\beta_{yyy}$	-423.2585
$\beta_{xxz}$	-94.3689
$\beta_{xyz}$	52.0453
$\beta_{yyz}$	44.7698
$\beta_{xzz}$	33.6780
$\beta_{yzz}$	68.7228
$\beta_{zzz}$	68.7228
$\beta_{total(esu)}$	$6.15906 \times 10^{-30}$

## 5. Result and discussion

### 5.1. Geometric structure

The optimized GPS monomer hops and dimer hops worked out by ab initio HF and DFT (B3LYP) level through the basis set of 6-31 G (d, p) are listed in (Table 3a) respectively and shown in figure 5 and 6. The bond lengths of C-N is 1.469Å for the amine group and 1.472Å for aziridine group the optimized single bond length range of C-N were 1.4504 – 1.465 Å by B3LYP/6-31G (d, p) method. As soon as the addition of the substituent groups (hydroxyl group) roots slight disparities between them. The length of the bond of C7-C10 for the title fragment has been 1.5161 Å and 1.5237 Å, formed by HF and abinitio DFT theory, on a base set of 6-31 G (d, p). Several authors [20] have clarified the increase in bond length of the C-H bond as a result of a change in carbon atom charge allocation. The exchange may be of the form of withdrawal (F, Cl, Br, N, etc.) or an electron form of donation. The strongly attracted carbon atom of the hydrogen cloud's valence electrons increases the constant CH force and reduces the bond length by the introduction of a number of substitutes. The opposite is valid for electron donor groups substitution. The real shift in the length of the C- H bond will be determined by the combined effect of mensomer-inductive contact and the electrical dipole field of the polar replacement. In this analysis, the bonding lengths of 1.0832, 1.0882 and 1.0798 Å have been measured as C2-H3, C7-H8, C7-H9, and C7-H10, but the length of the bond of C2-H4 is increased by about 0.01 Å because of phosphorous atom contact in this GPS community.

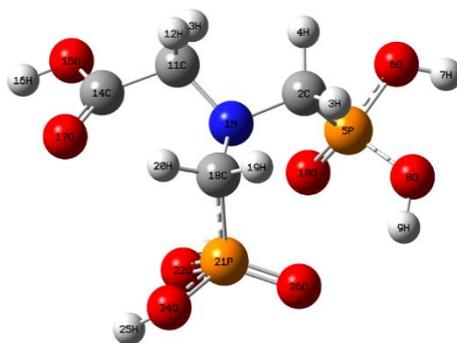


Fig. 5. Geometrical structure of Glyphosine.

The hops obtained using the B3LYP method with pole separated base sets are roughly the same as those tentatively observed. The defined bond length and bond angles are very close to the provisional values at all rates listed here. Interestingly enough, it has been drawn up using the HF method that both the P-O bond lengths between 1.4509 -1.5975, and 1.4791-1.6439 Å were formed using DFT Theory. The lengths of the P-O bond differ. The reversal between the single electron pair of the oxygen atom and the electron pair of the nitrogen atom can be due to this effect. For the monomer of the fragment named "DFT/ B3LYP/6-31G(d, p)" the bond longitude of O41-H42 and O15-H16 is 1.0042 Å. The fragments are stabilized by the intermolecular O – H — O bonds' endless chain. B3LYP/6-31 G (d, p) method is used to build computational dimer hops (intermolecular hydrogen bonding). From the (Table 3b), theoretical values, it can be found that most of the optimized bond lengths O41-H42--O17 and O15-H16---O43 are close to the bonding length value of 1,657 Å and the bond angle of corresponding value is 179.6 compared to the bond angles and length of B3LYP/6-31 G (d, p).

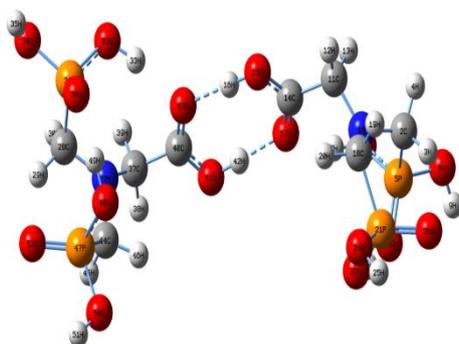


Fig. 6. Geometrical structure of dimer of Glyphosine.

Table 3a. Geometrical parameters optimized in (FA), bond length ( $\text{\AA}$ ), bond angle ( $^\circ$ ) and dihedral angle ( $^\circ$ ).

Parameters	HF 6-31G(d,p)	B3LYP 6-31G(d,p)	Parameters	HF 6-31G(d,p)	B3LYP 6-31G(d,p)
<b>Bond length (<math>\text{\AA}</math>)</b>					
N1-C2	1.4504	1.453	N1-C7-H8	112.2	110.5
N1-C7	1.4545	1.4643	N1-C7-H9	108.9	109.6
N1-C12	1.465	1.4696	N1-C7-C10	110.7	107.5
C2-H3	1.0832	1.0921	H8-C7-H9	109.4	108.6
C2-H4	1.0904	1.1006	H8-C7-C10	107.6	124.1
C2-P5	1.8086	1.8303	H9-C7-C10	108.0	111.5
P5-O6	1.4509	1.4791	C7-C10-O11	123.7	124.4
P5-O19	1.5975	1.6439	7C-C10-O23	112.3	112.1
P5-O21	1.5794	1.6117	O11-C10-O23	124.0	111.0
C7-H8	1.0882	1.0992	N1-C12-H13	112.3	109.1
C7-H9	1.0798	1.0899	N1-C12-H14	110.9	107.4
C7-C10	1.5161	1.5237	N1-C12-P15	108.7	108.7
C10-O11	1.182	1.2039	H13-C12-H14	107.2	108.4
C10-O23	1.3178	1.3425	H13-C12-P15	109.7	116.4
C12-H13	1.0849	1.0938	H14-C12-P15	107.9	102.1
C12-H14	1.0837	1.0935	C12-P15-O16	115.6	103.8
C12-P15	1.8099	1.835	C12-P15-O17	102.7	114.8
P15-O16	1.451	1.4816	C12-P15-O25	104.5	116.4
P15-O17	1.5811	1.6172	O16-P15-O17	114.7	101.2
P15-O25	1.5792	1.6104	O16-P15-O25	115.4	111.3
O17-H18	0.9438	0.9644	O17-P15-O25	102.2	109.4
O19-H20	0.9455	0.9667	P15-O17-H18	114.2	114.5
O21-H22	0.9443	0.9651	P5-O19-H20	112.2	107.2
O23-H24	0.9465	0.9696	P5-O21-H22	115.3	113.4
O25-H26	0.9486	0.9745	C10-O23-H24	108.9	110.5
<b>Bond angle (<math>^\circ</math>)</b>					
C2-N1-C7	114.2	114.7	P15-O25-H26	114.7	109.6
C2-N1C12	115.1	116.2	N1-C7-H8	112.2	107.5
			N1-C7-H9	108.9	108.6

Parameters	HF 6-31G(d,p)	B3LYP 6-31G(d,p)	Parameters	HF 6-31G(d,p)	B3LYP 6-31G(d,p)
<b>Bond length (Å<sup>o</sup>)</b>					
C7-N1-C12	113.9	114.6			
N1-C2-H3	110.0	110.2			
N1-C2-H4	113.3	113.7			
N1-C2-P5	114.9	114.4			
H3-C2-H4	106.7	106.7			
H3-C2-P5	103.9	104.1			
H4-C2-P5	107.4	107.0			
C2-P5-O6	115.6	116.5			
C2-P5-O19	104.4	104.8			
C2-P5-O21	104.1	101.7			
O6-P5-O19	111.9	111.7			
C6-P5-O21	114.5	117.0			
O19-P5-O21	105.1	103.5			

Table 3a. (Contd.)

Parameters	HF 6-31G(d,p)	B3LYP 6-31G(d,p)	Parameters	HF 6-31G(d,p)	B3LYP 6-31G(d,p)
<b>Dihedral angle (°)</b>					
C7-N1-C2-H3	155.2	153.9	O8-P5-O6-H7	-77.2	-125.1
C7-N1-C2-H4	35.9	34.2	O8-P5-O6-H7	173.8	2.0
C7-N1-C2-P5	-88.0	-89.2	H12-C11-C14-O15	12.2	70.0
C12-N1-C2-H3	20.8	16.5	H12-C11-C14-O17	-170.2	-14.4
C12-N1-C2-H4	-98.5	-103.2	H13-C11-C14-O15	-102.7	167.2
C12-N1-C2-P5	137.6	133.4	H13-C11-C14-O17	74.9	-132.8
C2-N1-C7-H8	-79.6	-78.0	C11-C14-O15-H16	177.1	48.7
C2-N1-C7-H9	41.7	43.2	O17-C14-O15-H16	-0.6	164.2
C2-N1-C7-C10	160.2	162.0	N1-C18-P21-O24	-163.8	-14.2
C12-N1-C7-H8	55.4	60.0	N1-C18-P21-O26	73.8	-71.9
C12-N1-C7-H9	176.6	-178.8	H19-C18-P21-O24	70.5	162.3
C12-N1-C7-C10	-64.8	-59.9	H19-C18-P21-O26	-52.0	57.4
C2-N1-C12-H13	14.4	10.5	H20-C18-P21-O24	-44.2	165.6
C2-N1-C12-H14	134.4	130.7	H20-C18-P21-O26	-166.7	39.9
C2-N1-C12-P15	-107.1	-109.9	N1-C18-O22-O23	19.2	-65.1
C7-N1-C12-H13	-120.1	-126.9	H19-C18-O22-O23	-114.7	49.2
C7-N1-C12-H14	-0.2	-6.7	H20-C18-O22-O23	127.6	-76.6
C7-N1-C12-P15	118.4	112.7	O24-P21-O22-O23	-173.5	178.5
N1-C2-P5-O6	-178.2	-175.6	O26-P21-O22-O23	-47.3	145.3
N1-C2-P5-O19	-54.7	-51.5	C18-P21-O24-H25	-147.2	18.5
N1-C2-P5-O21	55.3	56.0	O22-P21-O24-H25	103.0	-107.8

Parameters	HF 6-31G(d,p)	B3LYP 6-31G(d,p)	Parameters	HF 6-31G(d,p)	B3LYP 6- 31G(d,p)
<b>Dihedral angle (°)</b>					
H3-C2-P5-O6	-57.9	-55.3	O26-P21-O24-H25	-24.8	-68.0
H3-C2-P5-O19	65.5	68.8	H4-C2-P5-O10	174.2	-70.9
H3-C2-P5-O21	175.5	176.3	C2-P5-O6-H7	-77.2	-125.1
H4-C2-P5-O6	54.9	57.5	O8-P5-O6-H7	173.8	2.0
H4-C2-P5-O19	178.3	-178.4	O10-P5-O6-H7	49.4	128.7
H4-C2-P5-O21	-71.7	-70.9	C2-P5-O8-H9	142.6	178.1
C2-P5-O19-H20	-133.9	-125.1	O6-P5-O8-H9	-107.5	50.0
O6-P5-O19-H20	-8.1	2.0	N1-C11-C14-O15	137.2	-73.3
O21-P5-O19-H20	116.8	128.7	N1-C11-C14-O17	-45.2	108.5
C2-P5-O21-H22	157.5	178.1	H12-C11-C14-O15	12.2	-70.0
O6-P5-O21-H22	30.2	50.0	H12-C11-C14-O17	-170.2	-14.4
O19-P5-O21-H22	-93.0	-73.3	H13-C11-C14-O15	-102.7	167.2
N1-C7-C10-O11	110.6	108.5	H13-C11-C14-O17	74.9	-132.8
N1-C7-C10-O23	-69.4	-70.9	C11-C14-O15-H16	177.1	48.7

Table 3b. Intermolecular hydrogen bonding parameters based on B3LYP/6-31G (d,p) method.

O41-H42....O17	O41-H42	H42....O17	O43....O17	O41-H42....O17
	1.0042	1.657	3.283	179.6
O15— H16.....O43	O15—H16	H16.....O43	O23.....O39	O23—H22.....O39
	1.0042	1.656	3.283	179.6

### 5.2.1. Vibrational spectral analysis

The vibration study of GPS has been performed on the basis of methoxy, propenoic acid, hydroxy and phenyl ring mode characteristic vibrations. HF and density functional theory (B3LYP and BLYP), with a 6-31 G (d, p) base set, have been applied to theoretic calculations. The fragment of title is not planar and forms part of the category of C1 points. No corresponding distribution will exist for C1 symmetry. Three specific scaling factors were found in our research. HF/6-31G (d, p): 0.9026, BLYP/6-31G (d, p):0.9923 and B3LYP/6-31G (d, p): 0.9608[21,22] as shown in (Table 4).

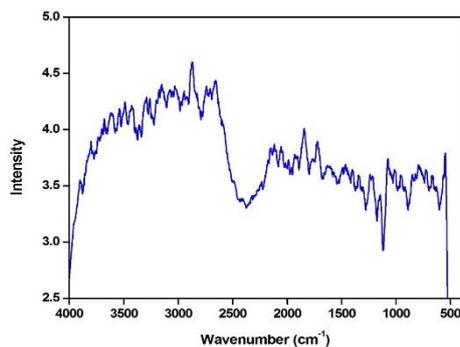


Fig. 1. Experimental FT IR spectrum of Glyphosine.

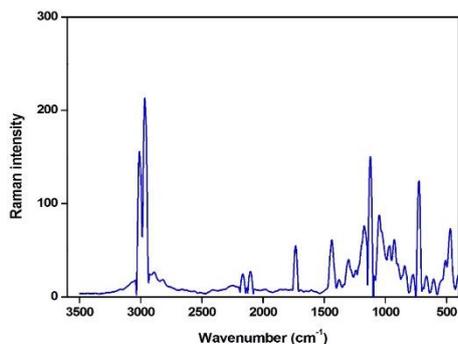


Fig. 2. Experimental FT Raman spectrum of Glyphosine.

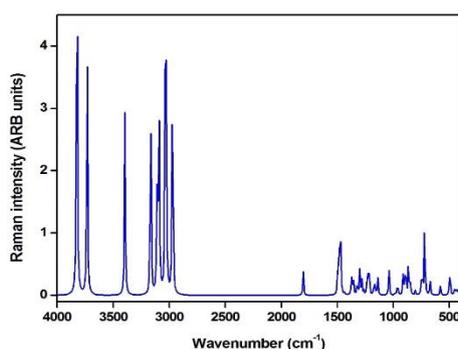


Fig. 3. Computed FT-IR spectra of Glyphosine.

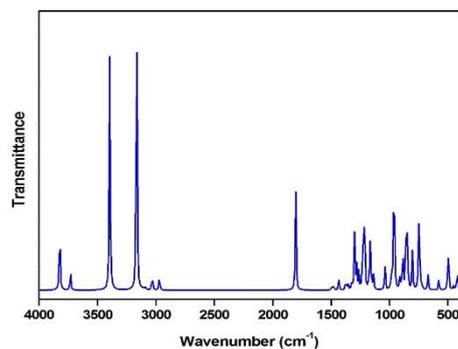


Fig. 4. Computed FT-Raman spectra of Glyphosine.

### 5.2.2. O-H vibrations

The O-H group produces three vibrations (spreading, winding and tearing out of the plane). The O-H group vibrations are most likely to be environmentally sensitive, displaying significant differences in the hydrogen-linked species' spectrum. In the area of about 3580-3650  $\text{cm}^{-1}$ , hydroxy stretching vibrations are normally [23] observed. The frequency of O-H vibration in the gas phase has been shown to be 3657  $\text{cm}^{-1}$  [24]. Similarly, the O-H stretching vibrations have been assigned to our FT-IR 3679, 3615 and 3490  $\text{cm}^{-1}$  band. Based on the literature data forecast, the harmonic approximation of the vibration intramolecular hydrogen bonds in the atomic group is a slight deviation. The in-ground frequency of the O-H bending in general is 1150-1250  $\text{cm}^{-1}$  and, unlike the expanding and out-of-ground bending frequencies of the hydrogen bond, it isn't affected much [25]. In the flat winding vibration, the weak band in FT-IR range is allocated to OH at 1147

$\text{cm}^{-1}$ . A good alignment with reported spectrum explains the theoretical measured  $1184\text{cm}^{-1}$  value by B3LYP process.

Bending mode of the O-H off-plan in the area  $517\text{-}710\text{ cm}^{-1}$  [26] is at a higher incidence than in free O-H in intermolecular and intramolecular associations. In our present study, an O-H out-of-plane bending vibration is allocated in the FT-IR range at  $551\text{ cm}^{-1}$ , the theoretically measured HF and B3LYP value also explains the same vibration form in the area  $684$  and  $580\text{ cm}^{-1}$  is assigned to O-H out-of- plane vibration.

### 5.2.3. C-H vibrations

The GPS fragment results in C-H stretching and C-H bending movements in the plane and C-H out of the plane. The C-H extending vibration is the key feature for identifying a C-heavily vibration in the area  $2850\text{-}3000\text{ cm}^{-1}$ , based on the literature analysis [27]. The anticipated C-H vibration is equal to the expanding modes (Mode No. 7-11) for the particle number. The Vibrational Mode nos (7-11), which are calculated with the  $2997, 2975, 2969, 2917, 2873$  and  $2862\text{ cm}^{-1}$ , are allocated to glyphosine stretching mode C-H computed with the FT-IR spectral recording at  $2646$ , the FT-Raman spectral at  $3012$ , and the FT-Raman spectral at  $2967\text{cm}^{-1}$ , respectively. In the  $2990\text{-}2861\text{ cm}^{-1}$  range, the same vibration degree of HF/6-31 G (d, p) calculated corresponds to the preliminary vibration.

In the area  $1350\text{-}1470\text{ cm}^{-1}$  is observed the C-H bending vibrations of alkanes and their plagiarism. The bands are sharp, but low to medium. A good deal of spectral compatibility with FT-IR is demonstrated by the  $1477\text{-}1463$  C-H bending vibrations measured HF and  $1435\text{-}1306\text{ cm}^{-1}$  by B3LYP/6-31 G (d) (d.p.). The same HF and B3LYP vibrations are also well consistent with provisional results. The theoretically developed C-H vibrations that stretch and bend are in good agreement with provisional values.

### 5.2.4. COOH vibrations

The extended band of carboxylic acids in the  $1705\text{-}1720\text{ cm}^{-1}$  range was generally observed. The band is induced by C = O vibration stretching. The double bond carbon oxygen is made by carbon and oxygen, because carbon and oxygen atoms have different electrical negativities. The single pair of oxygen electrons also decides about the character of the carbonyl group [28]. In such cases, two C = O are predicted, one of which is Raman active in the process (symmetric delaying of vibrations) and another of which is IR active in the segment (anti-symmetric delaying of caress) [29]. Due to hydrogen bonding in the next-door group – COOH – most carboxylic acids in their solid state form dimeric composition. In our current analysis, C = O stretching vibrations with strong concordance with the HF / B3LYP/6-31 G (d, p) value of the worked-out value at  $1812/756\text{ cm}^{-1}$  are assigned a powerful stripe observed in FT-IR spectrums of  $1722\text{ cm}^{-1}$  and FT-Raman spectrums of  $1734\text{ cm}^{-1}$ . In the region  $1395\text{-}1440\text{ cm}^{-1}$ , the characteristic carboxylic group C = O is expected to vibrate, according to whether the species are monomeric, dimeric and other hydrogen bound. In the theoretical calculations the C = O bending vibration is assigned to  $1437$  and  $1378\text{ cm}^{-1}$  by FT-Raman, according to the HF method, and with  $1367$  and  $1319\text{ cm}^{-1}$  by the B3LYP method.

### 5.2.5. HCH and HCN bending

Vibrations of HCH and HCN bend and separate vibrations of NH<sub>2</sub> and CH<sub>2</sub> groups from twisting and scissors [30]. HCH vible vibration waves with HF and the B3LYP system values  $1477, 1469, 1463\text{cm}^{-1}$  and  $1435, 1415, 1396\text{cm}^{-1}$  in the FT-IR range at  $1495, 1464$  and  $1394\text{ cm}^{-1}$  and  $1437\text{ cm}^{-1}$  in the FT-IR spectrum. The scaled HF and DFT / B3LYP methods for the HCN vibrations are formed in  $1359, 1343$  and  $1312\text{ cm}^{-1}$  and  $1311, 1283$  and  $1267\text{ cm}^{-1}$  wavenumbers.

### 5.2.6. Phosphorous group vibration

For Phosphorous functional group (P-OH) usually FT-IR region band is  $1040\text{-}930\text{ cm}^{-1}$  [31]. Detailed wave number of the named P-OH fragments for HF and  $1001.979, 970$  and  $956\text{ cm}^{-1}$  of the DFT / B3LYP process is  $1111, 1025, 1019$  and  $987\text{ cm}^{-1}$ . In our case, P-OH stretching mode in FT-Raman coincides theoretically with a high band with  $1049\text{ cm}^{-1}$  and an FT-IR with a high  $955\text{ cm}^{-1}$ .

The P = O-Stretching Mode for Phosphoramidate and Phosphate-1  $1200\text{-}1100\text{ cm}^{-1}$  and Plagiarist [32] is allocated in area  $1200\text{-}1275\text{ cm}^{-1}$ . In this case, the P = O stretch mode in FT-IR at  $1238\text{ cm}^{-1}$  is assigned as a band. The values for HF and DFT / B3LYP correspond with theoretical findings at  $1264\text{ cm}^{-1}, 1228\text{ cm}^{-1}$ . For lipids, P-O asymmetrical deformation mode is  $820\text{ cm}^{-1}$  [33].

The P-O asymmetrical stretch mode is available at 842  $\text{cm}^{-1}$  in the titled fragment, while the theoretically anticipated wave numerals for the HF and DFT / B3LYP system are 861-841 $\text{cm}^{-1}$  and 810-784  $\text{cm}^{-1}$ . In the waven numbers 420,403 and 388 $\text{cm}^{-1}$  for the HF-method and for 365,363 and 349  $\text{cm}^{-1}$  for the DFT / B3LYp method, the FT-Raman spectrum is assigned to P-O bending mode.

Table 4. Vibrational wave numbers obtained for Glyphosine at HF and B3LYP/6-31G (d, p) [harmonic frequency ( $\text{cm}^{-1}$ )].

Mode nos.	Experimental ( $\text{cm}^{-1}$ )		Wave number ( $\text{cm}^{-1}$ )				PED(%)
	FT-IR	FT-Raman	HF/6-31G(d,p)	IR <sub>int</sub>	B3LYP/6-31G(d,p)	IR <sub>int</sub>	
1	3679w		3682	216.36	3686	135.36	v-OH (100)
2	3615m		3680	204.02	3676	133.84	v-OH (99)
3			3645	139.92	3658	140.79	v-OH (99)
4			3512	527.64	3612	83.67	v-OH (100)
5	3490m		3373	724.17	3499	468.95	v-OH (100)
6		3012vs	2990	1.85	2997	2.15	v-CH (95)
7		2967vs	2960	13.76	2975	4.21	v-CH (14)+ v-CH (15)+ v-CH (68)
8	2946w		2951	8.68	2969	1.07	v-CH (27) + v-CH (44) + v-CH (27)
9			2919	31.15	2917	15.71	v-CH (58) + v-CH (41)
10	2868vs		2912	11.57	2873	56.83	v-CH (95)
11			2861	41.08	2862	31.52	v-CH (95)
12	1722vs	1734vs	1812	400.00	1756	277.51	v-OC (86)
13	1495w		1477	18.92	1435	9.36	T-HCCO (11) + $\beta$ -HCH (73)
14	1464m		1469	12.44	1415	18.96	$\beta$ -HCH (18) + $\beta$ -HCH (54)
15	1394m		1463	11.58	1396	0.84	$\beta$ -HCH (64) + $\beta$ -HCH (19)
16		1437s	1436	39.18	1367	28.52	v-OC (10) + v-CC (11) + $\omega$ -HCCO (21)
17		1378w	1381	9.47	1319	46.41	v-OC (13)
18			1359	14.16	1311	9.67	$\omega$ -HCNC (24) + $\omega$ -HCNC (27)
19			1343	52.87	1293	16.34	$\delta$ -HCNC (16) + T-HCNC (30) + $\gamma$ -HCH (13)
20		1300m	1312	46.55	1267	2.01	$\omega$ -HCNC (17)+ $\rho$ -HCNC (15) + $\gamma$ -HCN (31)
21			1288	133.60	1249	40.89	v-PO (14) + $\gamma$ -HCN (25) + $\beta$ -HCC (24)
22	1238m		1264	195.52	1228	164.68	v-PO (64) + $\gamma$ -HCN (12)

Table 4. ( Cond.)

Mode nos.	Experimental (cm <sup>-1</sup> )		Wave number (cm <sup>-1</sup> )				PED (%)
	FT-IR	FT-Raman	HF/6-31G(d,p)	IR <sub>int</sub>	B3LYP/6-31G(d,p)	IR <sub>int</sub>	
23		1243w	1242	206.27	1222	293.88	v-PO (64) + β - HCN (14)
24			1221	187.02	1204	29.11	β -HOC (11) + β -HCC (13)
25	1147w		1209	107.32	1154	126.24	v- OC (21) + β -HOC (24)
26		1173s	1167	247.94	1095	90.76	v-NC (21) + v-NC (20)
27			1152	143.25	1084	109.96	γ - HOP (73)
28		1123vs	1119	109.65	1081	14.80	v-NC (37) + β - HCN (10)
29		1049s	1111	61.81	1001	31.16	β -HOP (66)
30	955m		1025	85.25	979	50.10	β - HOP (81)
31			1019	70.30	970	64.85	γ -HOP (14) + ρ - HCCO (17)
32			987	14.23	956	44.44	γ -HOP (58)
33		965w	952	316.01	889	182.90	v-PO (15) + S-PO (22)
34			941	264.62	879	188.72	v-PO (34)
35		927m	905	46.23	857	51.32	v-OC (18)+ v-CC (28) + v- PO (12)
36			889	116.99	850	74.76	ω -HCNC (11) +v-PO(28)
37			884	33.95	832	45.43	v-NC (20) + v-NC (12)
38			861	280.57	810	328.28	v-PO (42)
39			854	64.85	806	37.61	v-PO (38)
40		842w	841	35.53	784	22.22	v- PO (42) + v-PO (14)
41	724w		748	42.56	720	52.15	γ -OCOC (27) + γ - OCO (10)
42		725vs	731	29.33	701	12.10	β -PCN (10) + v-PC(37)
43	666w		710	60.32	678	26.21	v-PO (17) + v-PC (34)+ v-PC(11)
44	551m		684	213.18	580	222.71	ω -HOPC (67)
45		664w	684	213.18	570	0.58	ω -HOPC (18) + β - OCO (42)
46			642	97.48	517	102.98	τ - HOCC (64)
47		605w	607	282.75	482	40.95	γ -OCOP (25)
48			561	41.51	457	44.53	β -OPO(12)+ β -OPO(13)+ β - OPO(14)

Table 4. (Cond.).

Mode nos.	Experimental (cm <sup>-1</sup> )		Wave number (cm <sup>-1</sup> )				PED (%)
	FT-IR	FT-Raman	HF/6-31G(d,p)	IR <sub>int</sub>	B3LYP/6-31G(d,p)	IR <sub>int</sub>	
49		508w	495	113.65	431	67.01	τ-HOCC (13) + β -CNC (25)
50			491	78.12	418	41.06	T-HOPC (17) + γ -OPO (22) + γ OPO (18)
51		467vs	452	19.31	412	14.92	ω -HCCO (11) + β -OCC (49)
52			437	4.46	392	22.16	β - OPO(11) + β - OPO (33)+ β - OPO (10)
53			420	103.87	376	5.42	γ -OPO (27) + ω -HOPC (10) + γ -OPO (27)
54		400s	403	43.30	365	59.35	γ -OPO (36) + γ -OPO (10)
55			388	21.13	363	36.34	γ -OPO (11) + γ -OPO(34) + γ -OPO (19)
56			375	22.48	349	120.32	γ -OCOP (11) + ω -HOPC (49)
57			333	2.62	306	6.33	γ -CNC(35)
58			331	9.15	279	3.10	ω -PCNC (10) + γ -OCOP(13)+ γ -CCCN (15)
59			296	8.35	267	33.22	γ -OCOP (11) + τ -HOPC (13)
60			277	5.87	224	18.67	γ -OCOP (17) + τ - HOPC(30)
61			241	13.59	215	13.58	γ -OCOP(17) + β -CNC (13)
62			223	25.09	192	33.66	β -OPO (18) + τ -HOPC (24)
63			181	90.79	157	5.11	γ -OCOP(10) + β -PCN (29)
64			175	17.55	144	3.74	γ -OCOP (21)+ γ -PCN (34)
65			168	86.33	141	43.39	ω - OPCN (24) + ω -OPCN (12) + ω -HOPC (23)
66			147	0.57	118	26.34	δ -PCNC (33)
67			144	3.27	92	23.04	δ -PCNC (21)+ ω -HOPC (22) + β -OPO (10)
68			133	0.70	80	3.66	δ -OCCN (31) + τ -CCNC (21) + ω -OPCN(16)
69			101	4.83	61	1.18	ω -OPCN(18)+ γ -CCCN (11) + β -CNC (10)
70			79	2.57	48	6.73	δ -OPCN (20) + ω -PCNC (24)
71			57	1.23	39	0.81	δ -CCNC (44) + ρ-PCNC(25)
72			42	1.91	34	0.70	δ -OCCN (19) + τ -OPCN (32)+ τ -OPCN (20)

IR<sub>int</sub>-IR intensity; Kmmol<sup>-1</sup>w-weak ; vw- very weak ; s-strong ; vs-very strong ; m-medium; v - stretching; β- in plane bending ; γ- out-of -plane bending ; ω - wagging ; t- twisting ; δ -scissoring ; τ- torsion.

## 6. HOMO and LUMO analysis

These orbitals are the bridge that is most closely associated with the energy of any pair of orbital fragments [34]. The intramolecular reassignment of the contributor to the assembly by a double-to-one-bond conjugated route will induce broad variations of molecular dipole and molecular polarization in order to enhance IR and Raman behavior simultaneously. The relative intensities from electron cloud movement through the conjugated frames of an electron donor to an electron acceptor group in Raman and in IR spectrums are also found in FT-IR glyphosine bands that have their complements in Raman. The preliminary spectrum is well defined by the initial calculations in conjugated systems which predict extraordinarily high Raman and infrared intensities in the same normal modes [35, 36]. The wave function study indicates that electron absorption corresponds to the transition from the ground into the first excited state and is determined primarily from the highest occupied molecular orbital (HOMO) to the lowest unoccupied molecular orbital (LUMO).

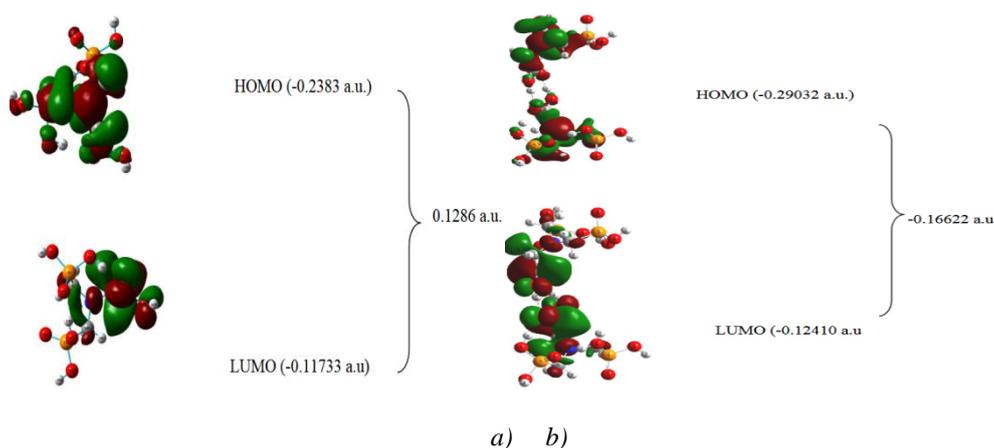


Fig. 7a. Compositions of the frontier molecular orbital for Glyphosine.  
 a) monomer atomic orbital  
 b) dimer atomic orbital

The energy divide of GPS from HOMO-LUMO has been worked out at level B3LYP/6-31 (d, p) and shows (fig. 7) that the difference in energy is indicative of the fragment's chemical activity (Table 5). LUMO's ability to get an electron is symbolic of HOMO's ability to contribute an electron. Titled fragments are formed in the DFT / B3LYP/6-31 (d, p) level at the dimer HOMO-LUMO energy gap as shown in (Fig. 8). The energy difference of the GPS fragment dimer is greater than the GPS fragment monomer. The self-compatible GPS energy field (SCF) is -1498.474 a.u. In fact, the lower energy difference between HOMO and LUMO describes the subsequent transition of charges within the fragment. The orbitals in the border region that display virtually degenerated energy.

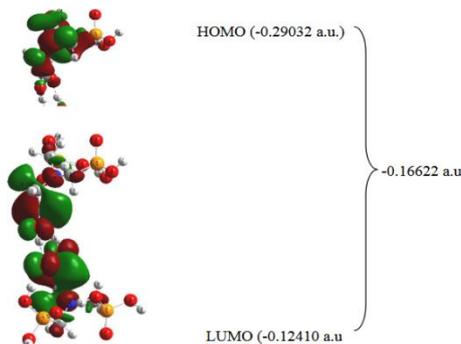


Fig. 8. The dimer atomic orbital compositions of the frontier molecular orbital for Glyphosine.

The Multiwfn study included complete state densities (TDOS), partial state density (PDOS) and partial overlap of states density (OPDOS) [37, 38]. The pictures of the state density show that energy of the functional group and the glyphosine energy element is within the range of -1.0 a.u. 6.0 a.u. A.u. In (Figure 9) showed. From the imagery of the homo-light-marked line overlapped with the density of states N, C-C and the hydroxyl group of the fragment named. The pictorial representation of the nitrogen and OH density distribution group is overlaid to the GPS fragments' homo-lump energy difference.

Table 5. HOMO-LUMO energy value calculated by HF/B3LYP/6- 31G (d, p) method for monomer and dimer of Glyphosine.

Parameters	Monomer		dimer
	B3LYP/6-31G(d,p)		B3LYP/6-31G(d,p)
HOMO (a.u)	-0.38646		-0.29032
LUMO(a.u)	-0.11733		-0.12410
HOMO-LUMO(a.u)	-0.26913		-0.16622

## 7. Potential energy surface scans

Figures 10a and 10b. 10a and 10b display the possible surface scan for the title fragment with the B3LYP/6-31 G (d, p) degree of theoretical approximation. The C11-C14-O15-H16 dihedral angle for GPS provides the appropriate co-ordinate for the elasticity of conformation of the fragments. The geometrical hops were all simultaneously loosened by B3LYP/6-31 G (d, p) measurement, while the torsional angle C11-C14-O15-H16 in 10o steps varied 10° to 360°. While for the dihedral angle of C11-C14-O15-H16, the maxima of the future energy curve at 190 ° (-1498.75529 Hatree), appears in 100° and 280°. The minimum total (-1498.74429 and -1498.74351 Hatree for the 280°) is probable due to the fact that methoxy and other hydrogen present in the fragment at this angle is not influenced by this hydroxy group. The C2-P5-O6-H9 dihedral angle was varied from 0° to 360° in 10° stage for named fragments. The prospective energy curve of two maxima and one minimum was observed. The C2 P5 O6-H9 average forward energy for scans is -1498.4814 for 40 degrees and -1498.4807 for 360 degrees and the lowest prospective scanning energy is -1498.4727 for 230 degrees. For dihedral angles (C11-C14-O25-H16 and C2-P5-O6-H9), the highest and lowest prospective scanning energy for the GPS fragments is present but prospective energy is not very varying because C-C and C-P will cause both dihedral angles.

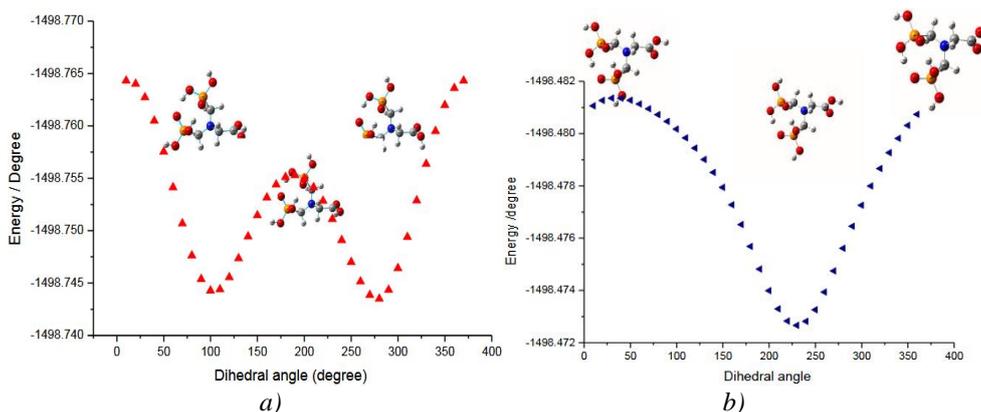


Fig. 10a. PES scan for dihedral angle  
 a) C11-C14-O15-H16 at b3lyp/6-31(d,p).  
 b) C2-P5-O6-H9 at b3lyp/6-31(d,p).

## 8. Thermodynamic properties

The theoretical method shown in Table 6 was used to perform statically thermodynamic functions such as a specific heat capacity, heat capacity and entropy changes with varying temperature from 100 to 1000 Kk. Based on the vibratory analysis, this thermodynamic hops can be observed as the temperature increases [39]. The corresponding correlation graph (Figure 11) of thermodynamic functions follows

$$S = 215.32 + 0.2973 T - 1.2 \times 10^{-4} T^2$$

$$C_p = 28.98 + 0.140 T - 7.6 \times 10^{-5} T^2$$

$$\Delta H = -2.523 + 0.049 T + 2.7 \times 10^{-5} T^2$$

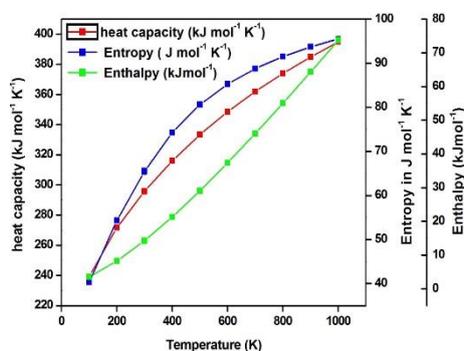


Fig. 11. Correlation graphic of heat capacity, entropy and enthalpy versus temperature for Glyphosine.

Table 6. Thermodynamic properties at different temperatures for Glyphosine.

T (K)	S (J/mol.K)	Cp (J/mol.K)	ΔH (kJ/mol)
100	311.48	87.59	5.35
200	394.69	158.69	17.77
298	469.51	218.43	36.35
300	470.86	219.46	36.76
400	541.09	269.2	61.29
500	605.51	308.01	90.23
600	664.45	338.16	122.6
700	718.44	362.13	157.66
800	768.12	381.74	194.88
900	814.05	398.2	233.9
1000	856.76	412.26	274.44

## 9. Natural bonding analysis

The bonding NBOs come from the orbital Lewis and the non-Lewis orbital antibonding NBOs. The complete Lewis orbital's are complemented by officially vacant non-Lewis orbital's with an idealized Lewis structure. The lower electronic load in Lewis orbital's shows good electron relocation effects. The Analysis of the NBO provides more information about the communication between empty and filled orbit which could improve intramolecular communication analysis. For each NBO donor NBO (i) and acceptor NBO (j) the Fock analysis perturbation theory is estimated for the second order: the delocation (2e-stabilization) "energy stabilization energy E(2) is estimated at [40,41].

$$E(2) = \Delta E_{ij} = q_i \frac{F(i,j)^2}{\epsilon_j - \epsilon_i} \quad (9)$$

The E(2) value for electron donor and electron acceptor communication is higher, and value is higher. An NBO study is performed by means of a named B3LYP fragment for the divulgation of electron density into fragments intramolecular, rehybridization and delocation [42, 43]. The Oxygen-Lone bonding pair and C-O antibonding pairs = 46.18 and 31.78 kcal / mol are recorded for the strongly stabilizing contact of  $\sigma_{O23} \rightarrow \sigma^*_{C10-O11}$  and  $\sigma_{O11} \rightarrow \sigma^*_{C10-O23}$ . An interaction between the O to C-O bond orbit forms the intramolecular hyper conjugate contact that results in the contact charge transfer that results in the stabilization within the fragment as well as in the results shown (Table 7).

Table 7. Second order perturbation theory analysis of Fock matrix in NBO basis for Glyphosine molecule.

Donor NBO (i)	TYPE	ED/e	Acceptor NBO (j)	Type	ED/e	E(2) kcal/mol	E(j)-E(i)	F(i,j)
P 5 - O 19	$\sigma$	1.98548	P5 - O21	$\sigma^*$	0.17439	3.34	1.05	0.055
C 7 - H 8	$\sigma$	1.98636	C10 - O23	$\sigma^*$	0.09654	3.53	0.89	0.051
C 7 - H 9	$\sigma$	1.97158	N 1 - C12	$\sigma^*$	0.02228	4.59	0.84	0.056
C 7 - C 10	$\sigma$	1.97849	N 1 - C2	$\sigma^*$	0.02371	3.37	0.98	0.051
C 12 - H 14	$\sigma$	1.98532	N 1 - C2	$\sigma^*$	0.02371	2.78	0.87	0.044
C 12 - P 15	$\sigma$	1.96847	P15 - O25	$\sigma^*$	0.16233	3.63	0.82	0.05
P 15 - O 25	$\sigma$	1.98713	P 15 - O 17 $\sigma^*$		0.16233	3.14	1.06	0.054
O 23 - H 24	$\sigma$	1.98364	C 7 - C10	$\sigma^*$	0.06850	4.09	1.09	0.061
O 6	$\pi$	1.99981	P 5	$\pi^*$	0.02618	3.32	19.55	0.229
O 11	$\pi$	1.99973	C10	$\pi^*$	0.02084	7.26	19.68	0.339
O 16	$\pi$	1.99973	P 15	$\pi^*$	0.05576	3.51	19.59	0.236
N 1	$\sigma$	1.99937	C 7 - H 8	$\sigma^*$	0.0694	7.66	0.68	0.066
O 6	$\sigma$	1.97993	P 5 - O 21	$\sigma^*$	0.17439	24.95	0.48	0.099
O 11	$\sigma$	1.97686	C10 - O 23	$\sigma^*$	0.09654	31.78	0.63	0.128
O 16	$\sigma$	1.97869	P15 - O17	$\sigma^*$	0.16233	25.48	0.48	0.1
O 17	$\sigma$	1.92561	P15 - O25	$\sigma^*$	0.16979	10.17	0.58	0.07
O 21	$\sigma$	1.96412	P 5 - O19	$\sigma^*$	0.18525	10.21	0.54	0.068
O 23	$\sigma$	1.91208	C10 - O11	$\sigma^*$	0.02263	46.18	0.34	0.113
O 25	$\sigma$	1.96178	P 15 - O 16 $\sigma^*$		0.07503	8.08	0.72	0.068
P 5 - O 21	$\pi$	1.98707	P 5	$\pi^*$	0.05798	12.14	0.53	0.211
P 15 - O 25	$\pi$	1.98713	P 15	$\pi^*$	0.05576	12.83	0.53	0.219

## 10. Mulliken charges

The application of quantum mechanical calculation to molecular system [44] is a critical atomic charge feature. The electrical charge of each atom is based upon the fragment's ability to bind. HF and B3LYP/6-31 G (d, p) method are used to evaluate the mulliken atomic charge of the GPS fragment. The GPS fragment's total

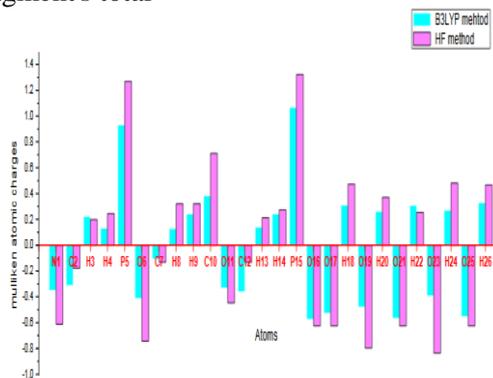


Fig. 12. Comparative of Mulliken's plot based on HF / B3LYP / 6-31G (d, p) method for Glyphosine.

Charges are (Fig. 12). It is evident from Table 8 that the P substitute of the fragment results in a higher electron density than the other alternating atoms. In the GPS fragment for HF and DFT / B3LYP process, the Mulliken atomic charge of the O atom gets more negative.

## 11. Conclusions

The frequency assignment for the GPS was carried out by FTIR and FT-Raman. On the theory level ab initio HF and DFT, the balancing geometries and harmonic GPS frequencies were defined and analyzed. The fact that the computations were made in the gaseous state with a single fragment, as opposed to the timely values recorded in the presence of intermolecular contact, may result from any observed divergence and the measured frequencies. There is a very small difference between the observed and the scaled values of wave numbers in many basic concepts.

Consequently, the tasks at DFT theoretical level tend to be accurate even with reasonable deviations from the provisional values. The FT-IR spectrum constructed theoretically coincides with the tentatively observed FT-IR spectrum. The DFT theory has drawn up two separate forward-looking energy scans for the hydroxy group called the fragment. The HF-method with 6-31 G (d, p) base set has worked out the electrical dipolar moments and first hyperpolarizability of the studied composite. The GPS fragment was analyzed on HOMO-LUMO energy and density distribution. The DFT / B3LYP system was used to perform the NBO analysis of Glyphosine. The energy of stabilization was examined by DFT method of natural bonding. This study shows that DFT / B3LYP calculations are a strong way of interpreting organic composite vibrations.

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